

# Pierce's Disease of Grape:

## IDENTIFICATION AND MANAGEMENT

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### Introduction

Pierce's disease (PD) of grape is caused by the bacterium *Xylella fastidiosa* (*Xf*) (Wilcox *et al.*, 2015). *Xf* is a xylem-limited bacterium. "Xylem-limited" means that *Xf* only inhabits the xylem elements within the host plant. Xylem elements are the tissues that transport water and nutrients from the roots up to the shoots and leaves. *Xf* limits water and nutrient translocation by persisting and expanding in the xylem elements and causing the plant to produce tyloses, blocking the "pipes" through which those resources can pass. If the *Xf* bacterium multiplies and expands enough, the result can be complete obstruction of water and nutrient translocation to aboveground vegetative tissues of the grapevine. Vines will consequently die within one to two years of PD diagnosis, which is well before the anticipated 20- to 25-year productive lifespan of a vineyard in the Eastern U.S. There are many biotic and abiotic pests that make wine grape cultivation challenging in the Southeastern U.S. PD can make it impossible to grow many wine grape cultivars in certain sub regions, as it kills them outright. PD is thus a great threat to the sustainability of commercial vineyards located in warm, coastal, and/or subtropical grape growing regions of the Southeastern U.S. as well as regions in Texas and California.

### Range and causal conditions

*Xf* is endemic to the Southeastern U.S. and is a threat to vine sustainability in sub regions (e.g. Piedmont, Coastal Plain, foothills, etc.) with mild winters. Pierce's disease can produce significant losses in vineyards from Florida to Virginia, but also in vineyards westward in Alabama, Texas, and Mississippi. PD is becoming increasingly common in susceptible cultivars (*V. vinifera*; popular hybrid bunch grapes such as Vidal blanc and Chambourcin) planted in northern Georgia and low-altitude North

Carolina and Virginia vineyards. The increasing PD prevalence in the Southeastern U.S. is likely a function of milder winter temperatures (Anas *et al.*, 2008; Lieth *et al.*, 2011). PD is generally less prevalent in vineyards located in northern, high altitude regions on the East Coast (Pennsylvania, New York) as well as inland regions that experience frigid winter temperatures, such as the Midwestern U.S. (Indiana, Michigan, Ohio). Pierce's disease has been identified, however, in Oklahoma and Missouri (Smith, 2013). It has generally been recommended that producers in Georgia not plant PD-susceptible vines at elevations below 1,300 feet above sea level, and rarely has PD been observed at elevations above 2,000 feet. However, PD has recently been observed to cause significant losses in vineyards at and above 1,300 feet; after warm winters, PD has even been confirmed in vineyards planted above 2,000 feet in northern Georgia, as well as in lower altitude vineyards in the North Carolina foothills.

### Symptoms and identification

As with several systemic grapevine diseases and nutrient imbalances, PD symptoms first become evident around veraison, the growth stage characterized by berry softening, sugar accumulation, and coloration (in red-fruited cultivars). Veraison is also the growth stage at which carbon resource allocation is shifted from the canopy vegetation to the ripening fruit; stressors become visually manifested in the canopy thereafter. Veraison typically occurs in mid- to late- July in many Georgia and Southeastern U.S. vineyards, but this will vary by cultivar, vintage, and growing region. Scouting for PD should begin in mid-July in those regions. Symptoms will occur



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earlier in drought years, but symptoms are generally most evident from late August through September. In grapevines, *Xf* causes four visual symptoms that together are strongly associated with PD. Leaf necrosis is observed on the margins of leaves (Figure 1A). Marginal leaf necrosis can be a common response to several vineyard issues; its presence alone should therefore not alarm growers. However, growers are encouraged to contact local Extension agents if the following symptoms are observed in tandem with marginal leaf necrosis. “Matchstick” is the term used to describe the symptom that occurs when only the leaf blade abscises and the petiole (leaf stem) remains attached to the shoot (Figure 1B). “Islands of green” describes the phenomenon observed when the shoot tissues harden off unevenly, resulting in green, vegetative “islands” within the brown, woody tissue (Figure 1C). As xylem flow is increasingly blocked by bacteria, infected plants will also form “raisins” as the fruit clusters dry out from lack of translocated water (Figure 1D).



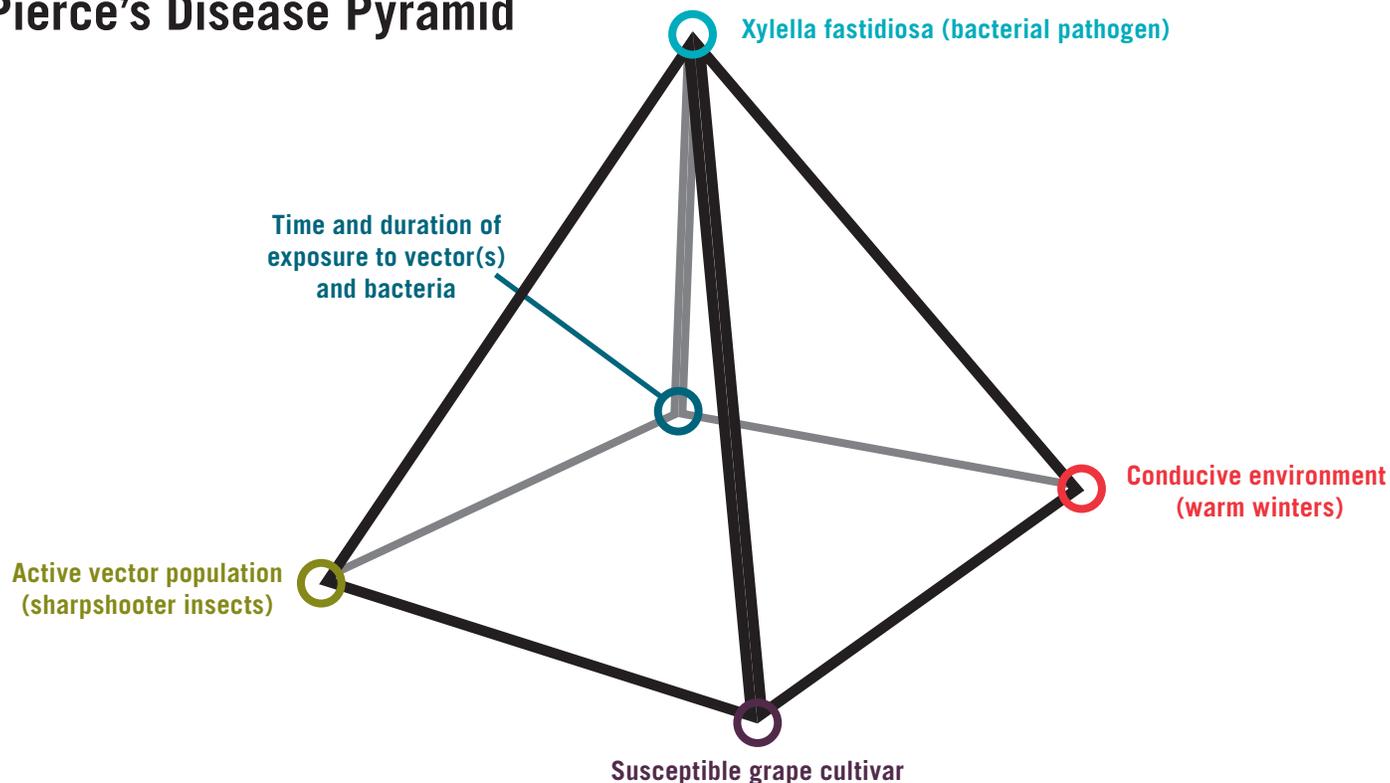
**FIGURE 1.** Pierce's disease symptoms include marginal leaf necrosis (A), leaf blade abscission from the attached petiole causing “match sticks” (B), “islands” of green tissue on primary grapevine shoots that are normally lignified and brown at this stage (C), and “raisins” (D).

## Management

The disease pyramid shows the necessary elements for grapevine PD to occur: a susceptible host, a conducive environment, a vector, an aggressive pathogen, and appropriate timing of vector feeding and bacterial ingress (Figure 2). A subfamily of leafhopper insects called “sharpshooters” are the significant means by which *Xf* is introduced into grapevines in the Southeastern U.S. Though pruning and root grafting are possible means of infection, these are considered to be of limited importance (Krell *et al.*, 2007). For this reason, without the presence of sharpshooter vectors, grapevine PD is unlikely to occur. The grapevine PD pyramid provides a good basis to develop a PD management plan in vineyards; exclusion or management of any of the five elements shown in the triangle below will result in successful, *proactive* grapevine PD management.

Annual dormant pruning has been anecdotally shown to aid in PD management in a commercial vineyard in Dahlenega, Georgia; failure to prune can exacerbate premature vine death due to PD. While there is no guarantee that pruning will remove all infected grapevine wood, annual winter pruning of 1-year-old canes can aid in PD management by removing first-year *Xf* infections (Varela *et al.*, 2001). Pruning may particularly aid in managing *Xf* infections that have occurred relatively later in the growing season, and thus have had a lesser chance of colonizing permanent vine parts before vines enter dormancy, relative to those infections that have occurred earlier in the season (Varela *et al.*, 2001). Indeed, vines infected by *Xf* earlier in the season (April and May) showed more severe PD symptoms and had less recovery rate relative to vines infected later in the season (June through August) (Feil *et al.*, 2002). While extremely impractical, cleaning tools with a 10% bleach solution can help reduce mechanical transmission of *Xf* to uninfected vines, particularly when hedging or trimming canopy shoots during the growing season (Smith, 2013). Once a vine is diagnosed to be PD-positive, the only 100% effective, *retroactive* management strategy is to rogue and remove the vine from the vineyard. PD symptoms are typically expressed later in the season, and identification may thus coincide with harvest time. If harvest activities preclude the immediate removal of infected vines, it is advised that infected vines be flagged so they can be easily identified for removal after the harvest is complete.

# Pierce's Disease Pyramid

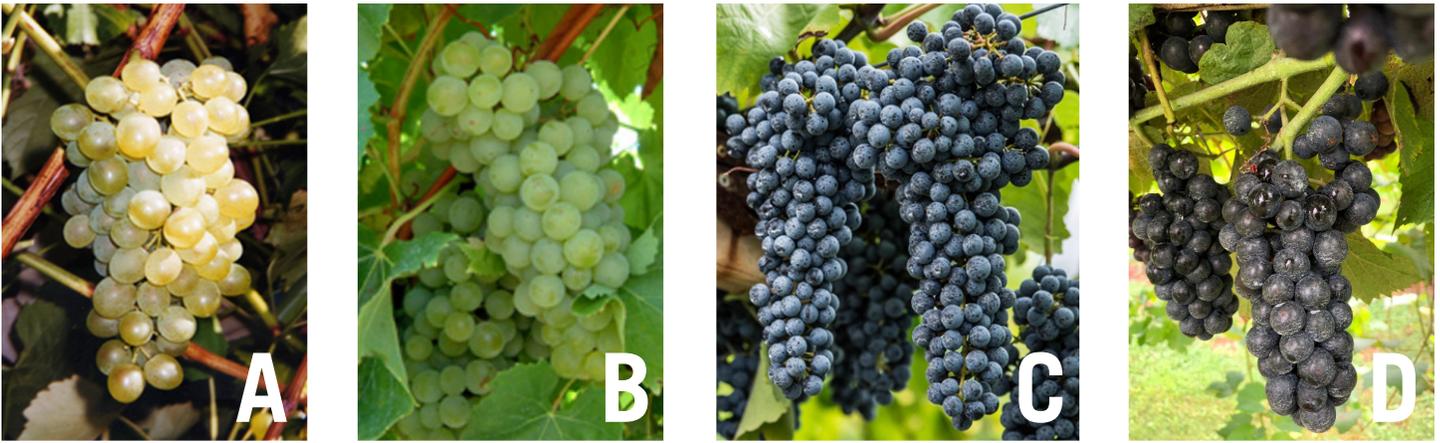


**FIGURE 2.** The five corners of the grapevine Pierce's disease (PD) pyramid. *Xylella fastidiosa*, a susceptible grape cultivar, a leafhopper vector, a conducive environment for pathogenicity (like that of the Southeast U.S.), and time and duration of exposure to the vector and pathogen all contribute to and are necessary for PD epidemic development.

The only way to “manage” a PD-conducive environment is to plant vineyards in northern regions or at elevations where winter temperatures are low enough to prevent the survival of *Xf* in the plant (Anas *et al.*, 2008; Feil and Purcell, 2001). Population numbers of key insect vectors may also be reduced in colder environments, but this impact of cold winters is likely secondary to reduced bacterial survival. Environments characterized by high and low PD threat can be found within the same state and are generally separated by a vague line of division that moves over time and between vintages. For example, the Texas Hill Country and north Georgia mountains are planted with PD-susceptible hybrid and *vinifera* cultivars, while the Texas Gulf Coast and Georgia Piedmont are planted with PD-tolerant cultivars.

Planting grapevine cultivars known to be tolerant to *Xf* is the best way to proactively manage PD in environments that are conducive to sustained *Xf* exposure. Tolerant cultivars are generally void of symptoms and their productivity is unaffected when planted in PD-conducive environments. Most muscadine cultivars and several hybrid wine grape

cultivars (e.g. ‘Blanc du Bois’ in Figure 3A, ‘Villard blanc’ in Figure 3B, ‘Lenoir’ in Figure 3C, ‘Norton’, etc.) tolerate *Xf* and therefore do not *often* show PD symptoms. While ‘Villard blanc’ is generally accepted as “PD-tolerant,” growers have reported visual PD symptoms in Villard blanc vineyards in the Southeastern U.S., and the same is true of Norton if grown under heavy PD pressure. Several T.V. Munson cultivars are also PD-tolerant (Kamas *et al.* 2010), the most popular of those being ‘Lomanto’ (Figure 3D). ‘Crimson Cabernet’, a privately bred, publicly available, red-berried cultivar developed from crosses between ‘Norton’ and ‘Cabernet Sauvignon’, has anecdotal PD-tolerance. High *V. vinifera*-parentage selections from Professor Andy Walker’s breeding program at University of California Davis tolerate PD and produce *vinifera*-like quality wines. White- and purple-skinned selections with  $\geq 94\%$  *vinifera* genetics are anticipated to be commercially released from Walker’s program over the next several years. Field trials are currently underway in Georgia to evaluate the growth and performance of ‘Crimson Cabernet’ and UC Davis selections.



**FIGURE 3.** Popular wine grape cultivars that are generally proven to be PD-tolerant and produce acceptable consumer wine quality, including 'Blanc du Bois' (A), 'Villard blanc' (B), 'Lenoir' (C), and 'Lomanto' (D). 'Blanc du Bois' and 'Villard blanc' photos courtesy of Fritz Westover; 'Lomanto' and 'Lenoir' photos courtesy of Joey Muller.

The PD-causing *Xf* is vectored and transmitted by xylem-feeding sharpshooters (Cicadellidae: Cicadellinae) and spittlebugs (Cercopidae) which feed on green, vegetative shoots of the grapevine. Many xylem-feeding leafhoppers, such as the broad-headed sharpshooter (*Oncometopia orbona*), versute sharpshooter (*Graphocephala versuta*), and the red-banded leafhopper (*Graphocephala coccinea*), are found in the Southeastern U.S. and can transmit the *Xf* bacterium (Krell *et al.*, 2007; Myers *et al.*, 2007; Redak *et al.*, 2004). The versute sharpshooter is commonly found in northern Georgia (Figure 4). The glassy-winged sharpshooter, *Homalodisca vitripennis* (Germar) (Hemiptera: Cicadellidae), is one of the most important vectors of *Xf* in the Southeast (Figure 4). This large, 0.5-inch-long sharpshooter is native to the Southeast, and its bulbous head and thorax (middle body segment) are dark brown with yellow dots. Its face, eyes, and legs are yellow-orange with dark markings. The glassy-winged sharpshooter has translucent, smoky brown, and membranous wings that cover its abdomen, which is white with black markings. A publication by Varela *et al.*, (2001) provides a thorough review of *Xf* insect vectors.

Sharpshooters feed and reproduce on a wide range of plants in a variety of habitats and prefer succulent over lignified growth, so grapevines are ideal sharpshooter habitats because they produce copious amounts of succulent vegetative growth. The glassy-winged sharpshooter is a strong flier relative to other sharpshooters, so it can disperse across the landscape and easily move into the vineyard and behind vineyard edges that are adjacent to forested habitats. With its high propensity for movement and its dark coloration, the glassy-winged sharpshooter is very inconspicuous

in the wild. Additionally, it often hides by moving to the other side of the cane when it detects movement or is disturbed. Thus, visually inspecting vines for glassy-winged sharpshooters can be tricky and ineffective, so using yellow sticky traps is a better tactic. The best time to start trapping for sharpshooters is at budbreak. Monitoring glassy-winged sharpshooters at this time will help determine activity and movement of sharpshooters into the vineyard from surrounding habitats. Use large, double-sided yellow sticky traps, no smaller than 4 by 7 inches, that contain plenty of sticky material (Varela *et al.*, 2001). Hang traps every 150 feet at canopy height throughout each block (area of contiguous rows of vines). Hang traps at the interface of the vineyard and all adjacent habitats, particularly forested and riparian borders. Check traps weekly and remove the insects from traps after counting them. Replace the yellow sticky traps at least once a month or when they are otherwise rendered inefficient at trapping and permitting efficient insect counting and identification. Continue monitoring for the glassy-winged sharpshooter throughout the season until daytime high temperatures remain below 65 °F (Varela, 2001).

The management of sharpshooters has been shown to suppress PD epidemics. One way to manage sharpshooters is to manage the habitat in which they feed and breed. Such attempts may prove futile because sharpshooters have a very wide range of natural and ornamental plant hosts. It is likely that succulent, green plant hosts are more ubiquitous in the Southeastern U.S. than in arid regions like California, where green vegetation is generally confined to irrigated and managed land. Aggressive vineyard weed and alleyway management may therefore reduce sharpshooter

breeding and feeding habitat, but several other problems related to the maintenance of soil structure and erosion prevention arise if the vineyard floor is barren of grasses. An indirect way of managing *Xf*-positive vectors would be to limit the number of *Xf*-harboring plants near vineyards. The difficulty lies in determining the plants that are worth managing as (1) many plants that harbor *Xf* are asymptomatic; (2) *Xf* multiplies and spreads to different extents across species and results in some plants being “better” inoculum sources than others; and (3) insect vectors may not feed on all *Xf*-positive plants (Varela *et al.*, 2001). Application of insecticides, particularly systemic insecticides, from the time of vector detection in the spring is advised (Krewer *et al.*, 2002). Glassy-winged sharpshooters often feed on the base rather than on the tips of canes, as well as on 2-year-old wood. Thus, early-season through mid-season management is particularly important, since early infections are more likely to lead to movement of bacteria to the cordon and trunk, where bacteria will not be removed through pruning practices (Feil *et al.*, 2003). UGA Extension Circular 1151 (Hickey *et al.*, 2018), “Viticulture Management,” is a poster offering a visual reminder and reference for sharpshooter scouting and management throughout the seasonal grapevine growth stages.



FIGURE 4. Glassy-winged sharpshooter (top) and versute sharpshooter (bottom).

## Testing

Grapevine tissues can be tested for PD infection using the polymerase chain reaction (PCR) method and enzyme-linked immunosorbent assay (ELISA). Using clean pruning shears, several petioles should be removed from vines showing PD symptoms. Many north Georgia vineyard owners can have their petioles tested for PD infection by bringing them to the Lumpkin or White County Extension offices. For a fee, the Plant Molecular Diagnostic Lab at the UGA Tifton campus can also test in-state or out-of-state samples. Those outside of Georgia should initially contact their local Extension agent or statewide small fruit or vineyard specialist for information about diagnosing PD on symptomatic vines, as other avenues for testing may be available.

## Summary

Pierce’s disease (PD) may be the greatest threat to the growth and sustainability of wine grape industries in the Southeastern U.S. The first step to managing grapevine PD is understanding the threat of PD as dictated by the region, or sub-region, in which vines will be planted. It is highly advised that PD-tolerant cultivars be planted if a vineyard will be established in a region of high PD-threat. Growers should understand that there is a risk of planting *Vitis vinifera* and other PD-intolerant cultivars in several Southeastern U.S regions, including the mountain regions of northern Georgia and piedmont regions in North Carolina. If PD-intolerant cultivars are planted, leafhopper vectors should be intensively scouted for and managed, and PD-infected vines should be immediately rogued out of the vineyard.

## References:

- Anas, O., Harrison, U. J., Brannen, P. M., & Sutton, T. B. (2008). The Effect of Warming Winter Temperatures on the Severity of Pierce's Disease in the Appalachian Mountains and Piedmont of the Southeastern United States. *Plant Health Progress*, 9(1), 13. doi:10.1094/php-2008-0718-01-rs
- Feil, H., & Purcell, A. H. (2001). Temperature-Dependent Growth and Survival of *Xylella fastidiosa* in Vitro and in Potted Grapevines. *Plant Disease*, 85(12), 1230-1234. doi:10.1094/pdis.2001.85.12.1230
- Feil, H., Feil, W. S., & Purcell, A. H. (2003). Effects of Date of Inoculation on the Within-Plant Movement of *Xylella fastidiosa* and Persistence of Pierce's Disease Within Field Grapevines. *Phytopathology*, 93(2), 244-251. doi:10.1094/phyto.2003.93.2.244
- Hickey, C. C., Blaauw, B., Brannen, P., Pfeifer D., Nita, M., Hoffmann, M. (2018). Viticulture Management. University of Georgia Extension Circular 1151. Retrieved from <https://extension.uga.edu/publications/detail.html?number=C1151>
- Kamas, J., Stein, L., & Nesbitt, M. (2010). Pierce's Disease-Tolerant Grapes. Texas A&M AgriLife Extension Publication. Retrieved from <https://aggie-horticulture.tamu.edu/fruit-nut/files/2010/10/pd-grapes.pdf>
- Krell, R. K., Boyd, E. A., Nay, J. E., Park, Y., & Perring, T. M. (2007). Mechanical and insect transmission of *Xylella fastidiosa* to *Vitis vinifera*. *Am J Enol Vitic.* 58:211-216.
- Krewer, G., Dutcher, J. D., & Chang, C. J. (2002). Imidacloprid Insecticide Slows Development of Pierce's Disease in Bunch Grapes. *Journal of Entomological Science*, 37(1), 101-112. doi:10.18474/0749-8004-37.1.101
- Lieth, J. H., Meyer, M. M., Yeo, K., & Kirkpatrick, B. C. (2011). Modeling Cold Curing of Pierce's Disease in *Vitis vinifera* 'Pinot Noir' and 'Cabernet Sauvignon' Grapevines in California. *Phytopathology*, 101(12), 1492-1500. doi:10.1094/phyto-08-10-0207
- Myers, A. L., Sutton, T. B., Abad, J. A., & Kennedy, G. G. (2007). Pierce's Disease of Grapevines: Identification of the Primary Vectors in North Carolina. *Phytopathology*, 97(11), 1440-1450. doi:10.1094/phyto-97-11-1440
- Redak, R. A., A. H. Purcell, J.R.S. Lopes, M. J. Blua, R. F. Mizell III, and P. C. Anderson. 2004. The biology of xylem fluid-feeding insect vectors of *Xylella fastidiosa* and their relation to disease epidemiology. *Annual Review of Entomology* 49: 243–270.
- Smith, D. 2013. Pierce's Disease. eXtension online article, February 28, 2013. Retrieved from: <https://articles.extension.org/pages/33077/pierces-disease>
- Varela, L.G., Smith, R.J., and Phillips, P.A. 2001. Pierce's Disease. University of California Davis, UC ANR Publication 21600. Retrieved from <http://iv.ucdavis.edu/files/24470.pdf>
- Wilcox, W.F., Gubler, W.D. and Uyemoto, J.K. 2015. Compendium of grape diseases, disorders, and pests. Second edition. St. Paul, MN: APS Press.

## Further reading:

- Kamas, J. Pierce's Disease Overview & Management Guide: A Resource for Grape Growers in Texas and Other Eastern U.S. Growing Regions. Texas A&M AgriLife Extension Publication. Retrieved from <https://aggie-horticulture.tamu.edu/fruit-nut/files/2010/10/Texas-Grape-Growers-PD-Management-Guide.pdf>
- Varela, L.G., Smith, R.J., and Phillips, P.A. 2001. Pierce's Disease. University of California Davis, UC ANR Publication 21600. Retrieved from <http://iv.ucdavis.edu/files/24470.pdf>

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